

Do Plants Have a Rapid Block to Polyspermy?

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Methods/Results Why TMEM16A? A table of expression values (RPM) for TMEM16A, EC1, and Genetically encoded membrane potential sensors are used to quantify the change in membrane potential by measuring the change in fluorescence. DD65 shows significant expression of TMEM16A localized in egg cell replicates. Mean Central Cell Expression Mean Egg Cell Expression Mean Synergid Cell Expression Gene AT1G73020 (TMEM16A) 0.1 AT1G76750 (EC1) 4738.6 AT3G10890 (DD65) CRISPR mutagenesis of TMEM16A has produced mutants with a deletion of the entire TMEM16A gene. Early PCR genotype analysis suggests the presence of possible homozygous mutants. 8252bp Guide Guide TMEM16A WT — Three possibly 1840bp obtained CRISPR Mu Sequencing results from the three mutants confirm the deletion of TMEM16A **Future Directions** Egg Cell Expressed Analysis of GFP fluorescence in an ovules containing ArcLight expressed in the egg cell **1.** Test for a plasma membrane potential change using the ArcLight sensor in TMEM16A CRISPR mutants 2. Test the protein localization of TMEM16A by tagging the C terminus with GFP **3.** Test the central cell plasma membrane for depolarization upon sperm fusion Conclusions EC1 ArcLight GFP Fluorescence (No Sperm The results from monitoring GFP fluorescence on the egg cell using the ArcLight sensor suggest that there is a depolarization event that occurs upon sperm fusion with 120 100 the egg cell. Depolarization across the plasma membrane of a female gamete is preliminary evidence for a rapid block to polyspermy in flowering plants. Analysis of membrane depolarization in TMEM16 CRISPR mutants should yield interesting results as TMEM16A is a conserved chloride ion channel implicated in the rapid block to polyspermy in animals. Frames (1 frame/10 seconds) EC1 Arclight GFP Fluorescence with Sperm Fusion The membrane depolarization seen in panel B can be seen as a sharp change in GFP fluorescence measured by the ArcLight sensor construct. Frames (1 frame/10 seconds)

